

1 **Skin advanced glycation end products as biomarkers of photosensitivity in**
2 **schizophrenia**

3

4 **Running title:** Carbonyl stress and photosensitivity

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7

1 **ABSTRACT**

2 **Objectives:** Photosensitivity to ultraviolet A (UVA) radiation from sunlight is an
3 important side effect of treatment with antipsychotic agents. However, the
4 pathophysiology of drug-induced photosensitivity remains unclear. Recent studies
5 demonstrated the accumulation of advanced glycation end products (AGEs), annotated
6 as carbonyl stress, to be associated with the pathophysiology of schizophrenia. In this
7 study, we investigated the relationship among skin AGE levels, minimal response dose
8 (MRD) with UVA for photosensitivity and the daily dose of antipsychotic agents in
9 patients with schizophrenia and healthy controls.

10 **Methods:** We enrolled 14 patients with schizophrenia and 14 healthy controls.
11 Measurement of skin AGE levels was conducted with AGE scanner, a fluorometric
12 method for assaying skin AGE levels. Measurement of MRD was conducted with UV
13 irradiation device.

14 **Results:** Skin AGE levels and MRD at 24, 48 and 72 h in patients with schizophrenia
15 showed a higher tendency for photosensitivity than in the controls, but the difference
16 was statistically insignificant. Multiple linear regression analysis using skin AGE levels
17 failed to show any influence of independent variables. MRD did not affect skin AGE
18 levels.

1 **Conclusions:** Photosensitivity to UVA in patients with schizophrenia receiving
2 treatment with antipsychotic agents might not be affected by skin AGE levels.

3

4 **Keywords:** schizophrenia, advanced glycation end products, carbonyl stress,
5 photosensitivity

1 1 | INTRODUCTION

2 Researchers have long sought validated, reproducible, sensitive and specific biomarkers
3 of schizophrenia. Previous studies have demonstrated significant alterations in the levels
4 of peripheral biomarkers, such as serum amino acids and inflammation markers levels
5 of schizophrenia (Chan et al., 2015; Nishimon et al., 2017; Ohnuma and Arai, 2011;
6 Ohnuma et al., 2008). While the majority of previous studies have demonstrated the
7 potential of diagnostic or prognostic biomarkers, few cross-sectional observational
8 studies have also examined the safety or toxicity of biomarkers for treatment with
9 antipsychotic agents (Lai et al., 2016; Tomasik et al., 2016). The accumulation of
10 advanced glycation end products (AGEs) associated with glycation stress is known as
11 carbonyl stress, and several recent studies have demonstrated an association between the
12 pathophysiology of schizophrenia and carbonyl stress (Arai et al., 2010; Katsuta et al.,
13 2014; Kouidrat et al., 2013; Miyashita et al., 2013; Takeda et al., 2015). Previously, we
14 reported that high serum levels of pentosidine, a marker of carbonyl stress, were
15 associated with high doses of antipsychotic agents and the duration of polypharmacy
16 treatment with antipsychotic agents in acute-stage schizophrenia (Katsuta et al., 2014;
17 Sannohe et al., 2017). Other researchers demonstrated possible correlation between
18 AGE levels and the duration of antipsychotic treatment as well as antipsychotic dose

1 (Hagen et al., 2017). Serum pentosidine levels have not been associated with the
2 severity of the symptoms of schizophrenia (Katsuta et al., 2014; Sannohe et al., 2017).

3 A photosensitive reaction, manifesting as a skin rash, is one of the side effects of
4 antipsychotic drug-induced toxicity. Photosensitivity is believed to reflect a disturbed
5 immune function after exposure of skin to sunlight and after the administration of not
6 only typical antipsychotics, including phenothiazine (Wolnicka-Glubisz et al., 2005) but
7 also current main stream of pharmacotherapy with atypical antipsychotics, including
8 clozapine, alanapine and aripiprazole (Al-Aojan and Al-Khalifah, 2018; Gregoriou et al.,
9 2008). In addition, some cases transferred to persistent light reaction with cross-reaction
10 mechanism, although the causal substance (antipsychotics) has been eliminated
11 (Amblard et al., 1982; Barbaud et al., 2001). The main mechanism of photosensitivity
12 was considered to be immunoreaction, especially delayed-type allergy. Photohaptens are
13 the main causative substances in first sensitization of photosensitivity and bind covalently
14 to protein under exposure to UVA via Langerhans cells photomodified with a
15 photohaptent, which can stimulate immune T cells; then these mechanisms were elicited
16 by UVA irradiation (Tokura, 2000). Finally, these reactions cause skin inflammation that
17 was also caused by accumulation of AGEs, which was caused by antipsychotics dose and
18 considered as one of the pathophysiologic mechanisms of schizophrenia (Ohnuma et al.,

1 2018). Thus, the accumulation of skin AGEs also might be involved in the
2 pathophysiology underlying drug-induced photosensitivity. Indeed, the accumulation of
3 AGEs in skin has been shown to be involved in the pathophysiology of some
4 dermatological diseases, such as solar elastosis (Yoshinaga et al., 2012), prurigo
5 nodularis in dialysis patients (Dyer et al., 1993; Meng et al., 2001) and perforating
6 dermatosis (Fujimoto and Tajima, 2004; Seite et al., 1998).

7 In the current study, we investigated the relationship between photosensitivity and
8 possible causative factors, including skin AGE levels, minimal response dose (MRD),
9 daily antipsychotic doses and serum pentosidine and pyridoxal levels in patients with
10 schizophrenia treated with antipsychotic agents and in healthy controls. We aimed to
11 investigate whether a high glycated stress state is associated with overdose of
12 antipsychotic drugs and the potential implication of using glycated stress as a biomarker
13 of photosensitivity in schizophrenia.

14

15 **2 | METHODS**

16

17 **2.1 | Subjects**

1 The Ethics Committee of the Juntendo University School of Medicine approved the
2 study protocol (15-052). All participants provided written informed consent prior to
3 participation. Fourteen stable outpatients diagnosed with schizophrenia and treated with
4 a fixed dose of antipsychotics for ≥ 30 days were enrolled in a cross-sectional
5 observational study at Juntendo University Schizophrenia Project (Ohnuma et al., 2008).
6 First-episode, drug-naïve patients were excluded from this study. All included patients
7 met the Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition (DSM-5)
8 criteria for schizophrenia (Association, 2013). Clinical symptoms were assessed using
9 the Brief Psychiatric Rating Scale (BPRS), with each item rated on a seven-point scale
10 as previously described (Katsuta et al., 2014). Three patients treated with
11 first-generation antipsychotics, ten treated with second-generation antipsychotic and one
12 treated with both types of antipsychotics (polypharmacy) were enrolled. Total
13 administered antipsychotics were converted to the chlorpromazine (CP) dose (Inada and
14 Inagaki, 2015). In addition, 14 healthy controls without current or a history of psychosis
15 according to the Japanese latest version of Structured Clinical Interview for DSM-IV
16 were recruited through the website of the Juntendo Hospital. The inclusion criteria in
17 this study were as follows: 1) age between 20 and 60 years to exclude ageing-related
18 effects; 2) no photosensitivity caused by factors such as porphyria or xeroderma

1 pigmentosum syndrome; 3) no polypharmacy, i.e. no prescription history of >3
2 antipsychotic drugs; 4) no history of diabetes, atopic dermatitis or kidney dysfunction;
3 5) no more than moderate obesity (i.e. BMI < 30 kg/m²); 6) no medication use resulting
4 in hyperesthesia optica and 7) no history of smoking or alcoholism. A schematic of the
5 study protocol is presented in Figure 1.

6

7 2.2 | Measurement of pentosidine

8 All blood samples were collected prior to breakfast to control for the influence of food
9 and exercise. The samples were immediately centrifuged at room temperature for 5 min
10 at 3500 rpm (2400 ×g), and the supernatant was frozen at −80 °C until further use. The
11 concentration of serum pentosidine was measured using a competitive enzyme-linked
12 immunosorbent assay kit (FSK Pentosidine, FUSHIMI Pharmaceutical Co., Ltd.,
13 Kagawa, Japan), as previously described. Briefly, pronase was added to the serum to
14 expose the protein-bound pentosidine molecules, and the samples were incubated at
15 55 °C for 1.5 h. After the stipulated reaction time, the mixtures were heated in boiling
16 water for 15 min for enzyme inactivation. An antibody against pentosidine and the
17 pre-treated sample or a standard pentosidine solution were added to each well and
18 incubated at 37 °C for 1 h. After washing, a peroxidase-labelled goat anti-rabbit IgG

1 polyclonal antibody was added and incubated at room temperature for 1 h. Subsequently,
2 a colour development reagent containing 3,3',5,5'-tetramethylbenzidine (TMB) was
3 added to each well. The reaction was stopped 10 min later by adding a TMB stop buffer.
4 Absorbance was measured within 10 min at 450 and 630 nm (main and reference
5 wavelength, respectively) (Katsuta et al., 2014).

6

7 2.3 | Measurement of pyridoxal

8 Serum pyridoxal levels were measured using high-performance liquid chromatography
9 (HPLC) on a Hitachi L-7000 series (Hitachi, Ibaraki, Japan). Then, 200 μ l aliquots of
10 the serum were added to 200 μ l citrate buffer solution and 75 μ l acid phosphatase
11 followed by incubation at 37 °C for 1 h. After completion of hydrolysis, 300 μ l of 20%
12 trichloroacetic acid was added to the mixture followed by centrifugation for 10 min at
13 12000 rpm (12000 \times g) at 4 °C. Subsequently, 400 μ l of the upper aqueous layer was
14 neutralised with 75 μ l sodium acetate solution and used as HPLC reagent. The HPLC
15 reagents were analysed with a Fluorescent Detector FP-2025 (Ex λ : 325 nm, Em λ : 395
16 nm; JASCO Corporation, Tokyo, Japan) and a Wakosil-II $_5$ C₁₈HG (4.6 ϕ \times 250 mm)
17 column (Wako Pure Chemical Industries, Osaka, Japan). Detailed information regarding
18 the HPLC conditions can be accessed in a previous study by (Katsuta et al., 2014).

1

2 2.4 | Measurements of skin AGE levels

3 Measurement of skin AGE levels was conducted with a TruAge Scanner™ (Morinda
4 Worldwide Inc.), which represents a fluorometric method for assaying AGE levels in
5 the skin. The forearm of the patients' dominant arm was placed on the scanner. A
6 prerequisite for this assay is that the skin in the area being measured must be healthy;
7 homogeneous; free of birthmarks, tattoos or excessive hair growth and without recent
8 exposure to skin creams or any substance possibly having fluorescent properties. All
9 measurements lasted approximately 15 s and was performed in triplicate after which the
10 results were presented as an average value (Hagen et al., 2017).

11

12 2.5 | Measurement of MRD

13 Measurement of MRD was conducted with UV irradiation device (UV109A, Herbert
14 Waldmann GmbH & Co., KG). We conducted a “preliminary study” with some healthy
15 volunteers, our preliminary data (unpublished) showed that MRD range of UVA in
16 Japanese healthy controls were 15–30 J/cm² with present methods. In addition, the
17 lowest MRD is known as 320kJ/m² for UVA (360nm), approximately 30 J/cm², and
18 appearance of erythema at 24 to 72 h with under 15 J/cm² UVA radiation means these

1 subjects assumed to have photosensitivity (Hawk et al., 2013). Thus, the maximum
2 irradiation time was defined as 30 J/cm² in this study. Eight patch holes (0% = 30J/cm²,
3 20%, 30%, 40%, 50%, 60%, 70% and 80%) were selected in a V strength rate cut-off
4 filter. First, we irradiated the skin on the patients' front arm with UVA light. After 24,
5 48 and 72 h, the exposed skin area was examined during a hospital visit
6 (<http://www.jove.com/video/50175>) (Heckman et al., 2013). Unfortunately, six patients
7 who underwent MRD at 24 h, one who underwent MRD at 48 h and two who
8 underwent MRD at 72 h were unable to visit the hospital owing to personal reasons.
9 Thus, the number of patients who underwent MRD at 24, 48 and 72 h was considered to
10 be 8, 12 and 13, respectively.

11

12 2.6 | Statistical analysis

13 All statistical analyses were conducted using SPSS version 22 (IBM Corp., Armonk,
14 NY). Chi-square tests were conducted to analyse the difference in terms of sex
15 distribution between the groups. A *p*-value < 0.05 was defined as the level of statistical
16 significance. Differences in terms of serum pentosidine, pyridoxal and skin AGE levels
17 and MRD between the unpaired groups were examined using a two-tailed Mann–
18 Whitney *U*-test for comparison between the two groups (schizophrenia vs. healthy

1 control). Correlation between the clinical features (e.g. age, BMI and daily CP
2 equivalent dose) and measured biomarkers was analysed using single correlation
3 analysis (Spearman's correlation test).

4 Multiple linear regression analysis included factors that potentially contributed to
5 significantly different skin AGE levels. These factors were selected on the basis of their
6 significant correlation with reported altered clinical characteristics such as BMI, age and
7 daily CP equivalent dose and were set as independent variables. Finally, stepwise
8 multiple regression analyses were performed for the potential significantly different skin
9 AGE levels as dependent variables and for the three above-mentioned clinical
10 variables as independent variables. The post-hoc power analysis ($1 - \beta$) were performed
11 with G*Power (<http://www.gpower.hhu.de/>) with the condition; $\alpha = 0.05$, effect size =
12 0.5 and each sample group size = 14.

13

14 **3 | RESULTS**

15

16 3.1 | Comparison of clinical variables

17 Clinical variables including the measured biomarker levels were compared between the
18 patients and healthy controls (Table 1). Sex ratio, age and BMI as well as serum

1 pentosidine did not differ between the groups (Table 1). However, the serum pyridoxal
2 levels in the patients were significantly lower than in the healthy controls (Table 1).
3 While skin AGE levels in the patients with schizophrenia showed a higher tendency
4 compared with the controls, they did not reach statistical significance. Furthermore, the
5 MRD at 24, 48 and 72 h showed higher tendencies in the patients with schizophrenia
6 than in the controls, but the difference was also statistically insignificant. Post-hoc
7 power analysis for this statistic showed $(1 - \beta) = 0.08$.
8 Single correlation analysis was performed among the carbonyl stress markers (serum
9 pyridoxal and pentosidine and skin AGEs), MRD (24, 48 and 72 h) and the three
10 clinical variables (age, BMI and daily CP equivalent dose) in patients with
11 schizophrenia (Supplementary Table 1). As expected, skin AGE levels showed a
12 significant correlation with age ($r = 0.477$, $p = 0.010$) using single correlation analysis,
13 but not with BMI ($r = 0.114$, $p = 0.562$) and daily CP equivalent dose (patients, $r =$
14 0.018 , $p = 0.952$). Other carbonyl stress markers did not show significant correlation
15 with any clinical variables (all $p > 0.05$). MRDs at 48 h ($r = 0.386$, $p = 0.047$) and 72 h
16 ($r = 0.394$, $p = 0.047$) showed a marginally significant correlation with BMI. In the
17 nature of things, three MRDs showed significant correlations with each other

1 (Supplementary Table 1). Post-hoc power analysis for this statistic showed $(1 - \beta) =$
2 0.07.

3

4 3.2 | Multiple linear regression analysis

5 Stepwise multiple linear regression analysis was performed in patients with
6 schizophrenia using skin AGE levels as a dependent variable. Age, BMI and CP
7 equivalent dose showed a significant association with skin AGE levels in the present
8 study (age) and previous studies (BMI and CP equivalent dose) (Sannohe et al., 2017;
9 Takeda et al., 2015) as independent variables. In this analysis, the three independent
10 variables were excluded in the first stage of the analysis in predicting skin AGE levels.
11 However, forced-entry multiple linear regression analysis was performed using the
12 same dependent and independent variables; however, no independent variables were
13 shown to affect skin AGE levels ($F = 0.61, p = 0.62$). The stepwise multiple linear
14 regression analysis was performed using MRD (24, 48 and 72 h) as a dependent
15 variable and age, BMI, CP equivalent dose and skin AGE levels as independent
16 variables. The results of these analyses showed that BMI was a significant variable for
17 the prediction of MRD for 72 h in the first stage of analysis ($F = 11.1, p = 0.009$),
18 whereas age, CP equivalent dose and skin AGE levels were not significant. The

1 predictive equation for calculating MRD at 72 h was as follows: $\text{MRD at 72 h (J/cm}^2\text{)} =$
2 $-6.6 + 0.98 \times \text{BMI}$. All independent variables were excluded in the first stage of the
3 analysis for predicting skin AGE levels for MRD at 24 and 48 h. Post-hoc power
4 analysis for this statistic showed $(1 - \beta) = 0.06$.

5

6

7 **4 | DISCUSSION**

8

9 In the present study, we investigated the relation between photosensitivity using MRD
10 and variable antipsychotic doses as the potential causative pathophysiological agents
11 underlying the accumulation of skin AGEs. The results that higher tendency of serum
12 pentosidine levels and significantly lower levels of serum pyridoxal in the patients with
13 schizophrenia than in the healthy controls were consistent with and confirmed previous
14 well-established results of studies that used serum biomarkers as indicators of carbonyl
15 stress in schizophrenia (Arai et al., 2010; Katsuta et al., 2014; Kouidrat et al., 2013;
16 Miyashita et al., 2013; Takeda et al., 2015). These results verified that the subjects
17 enrolled in this study possibly presented features of carbonyl stress in a population
18 suffering from schizophrenia. While skin AGE levels in the patients with schizophrenia

1 showed a higher tendency than in the healthy controls, they did not reach statistical
2 significance. Measurement of skin AGE levels with the TruAge Scanner reflected on
3 the levels of pentosidine, crossline (Obayashi et al., 1996) and pyrropyridine (Hayase et
4 al., 2008) as those exerting fluorescence and crosslinkage with collagen in the skin
5 (Meerwaldt et al., 2005). Although apparent measured AGEs were not specifically
6 restricted (Sell et al., 2018), skin AGE levels showed a clear correlation with the
7 concentration of skin pentosidine levels (Meerwaldt et al., 2005). The accumulation of
8 skin AGEs related to risk for further complications in diseases such as cardiovascular
9 disease, macular degenerative disease etc (Bos et al., 2011; Lutgers et al., 2009) could
10 prove useful as a surrogate marker of the status of carbonyl stress in the skin (Macasai et
11 al., 2013). Thus, the status of carbonyl stress in patients with schizophrenia might vary
12 depending on the affected tissues, e.g. accumulation in the skin or peripheral blood.

13 The current study has certain limitations, such as the small number of subjects (14
14 patients with schizophrenia) with low statistical power ($1-\beta$; from 0.06 to 0.08)
15 compared with a previous large serum AGE level-driven study (274 patients with
16 schizophrenia) (Sannohe et al., 2017). Further, all patients of the present study were
17 outpatients who showed milder symptoms and were prescribed lower daily doses
18 antipsychotic agents compared with previous studies that included patients suffering

1 with severe schizophrenia (Sannohe et al., 2017). Both factors, namely, mild disease
2 symptoms and low dosing of antipsychotics, may affect the levels of peripheral
3 carbonyl stress markers (Arai et al., 2010; Katsuta et al., 2014; Kouidrat et al., 2013;
4 Miyashita et al., 2013; Takeda et al., 2015). Indeed, skin AGE levels were significantly
5 elevated in patients with severe schizophrenia (Kouidrat et al., 2015). Furthermore, in
6 skin AGEs levels, elevated these showed relation with antipsychotics dose and its
7 accumulative exposure periods in patients with a recent onset of psychosis (Hagen et al.,
8 2017), as we also showed in serum study (Sannohe et al., 2017).

9 Unfortunately, this study failed to demonstrate any significant association
10 between daily antipsychotic drug dose and MRD with UVA on skin AGE levels.
11 Further, a control for the specific type of antipsychotic drugs was not accounted for in
12 this study. Among the 14 patients with schizophrenia, only 1 patient treated with
13 polypharmacy showed the most influenced peripheral AGE levels (Sannohe et al., 2017).
14 Although the results presented in this study are clinically relevant, they need to be
15 reproduced in studies that include larger sample sizes and patients with severe
16 schizophrenia treated with high doses of antipsychotic drugs. Patients treated with
17 antipsychotic drugs showed a minor tendency to exhibit lower photosensitivity to UVA.
18 Additionally, higher BMI might influence the MRD at 72 h to show a higher value (i.e.

1 lower photosensitivity to UVA) in patients with schizophrenia. Obesity has been
2 suggested to trigger a higher status of carbonyl stress (Miyata et al., 2005; Nangaku et
3 al., 2005). The current results demonstrated an inverse relation between BMI and MRD.
4 Overall, we conclude that photosensitivity to UVA in patients with schizophrenia was
5 not clinically apparent and was unaffected by skin AGE levels and related factors such
6 as low antipsychotic drug dose.

7 Further studies using non-invasive measurement of skin AGE levels on a larger
8 subject population, including those suffering from severe schizophrenia and especially
9 those with demonstrated history of photosensitive reaction to antipsychotic agents,
10 might be beneficial in clinical practice for the treatment of schizophrenia.

11

12 **CONFLICT OF INTEREST**

13 None to declare

14

15 **AUTHORS CONTRIBUTIONS**

16 Eriko Tani and Tohru Ohnuma designed the study protocol, recruited subjects, collected
17 clinical data, performed statistical analysis and wrote the manuscript. Yuto Takebayashi,
18 Narimasa Katsuta and Shohei Nishimon designed the study, recruited subjects and

1 collected clinical data. Hitoki Hirose, Ken Nakayama, Wanyi Mao, Mariko Nakadaira,
2 Narihiro Orimo, Hiroki Yamashita and Yasue Miki assisted in manuscript preparation.
3 Toshio Hasegawa, Etsuko Komiyama, Yasushi Suga and Shigaku Ikeda contributed to
4 study design and interpretation of analysed dermatological data. Heii Arai contributed
5 to the interpretation of all analysed data. All authors contributed to manuscript
6 preparation and approved the final version.

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1 **Table 1.** Comparison of clinical variables between the patients with schizophrenia and
 2 healthy controls

Variables	Patients with Schizophrenia	Healthy Controls	Statistical Test and <i>P</i> -value	
	(n = 14)	(n = 14)	<i>t</i>	<i>P</i> -value
Sex, M/F	3/11	8/6	3.743 (χ^2)	0.120
Age, mean (y)	42.6 ± 10.7	35.7 ± 9.6	-1.819	0.069
BMI (kg/m ²)	22.9 ± 4.39	21.8 ± 3.24	-0.965	0.335
DOI (y)	18.1 ± 11.7	NA	NA	NA
Total BPRS scores	24.5 ± 3.1	NA	NA	NA
CP equivalent dose (mg/day)	333.9 ± 234.9	NA	NA	NA
Serum pyridoxal (ng/ml)	10.65 ± 6.52	27.58 ± 38.51	-2.112	0.035
Serum pentosidine (ng/ml)	48.27 ± 14.65	45.94 ± 14.69	-0.291	0.771
Skin AGEs (AU)	246.45 ± 65.17	211.69 ± 31.96	-1.817	0.108
MRD 24 h (J/cm ²)	17.63 ± 8.40	13.93 ± 6.82	-0.997	0.319
MRD 48 h (J/cm ²)	14.54 ± 6.46	14.36 ± 6.98	-0.222	0.825
MRD 72 h (J/cm ²)	16.25 ± 6.45	14.79 ± 7.00	-0.316	0.752

Variable data are presented as mean ± standard deviation and range.

AGEs, advanced glycation end products; BMI, body mass index; BPRS, Brief Psychiatric Rating Scale; CP equivalent dose, chlorpromazine equivalent dose; DOI, duration of illness, MRD, minimal response dose

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1 **Figure legends**

2

3 **Figure 1 Inclusion criteria and measurement of minimal response dose (MRD) to**

4 **UVA**

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